Lest Update: 24/05/2019 Ecology: Coastal cliffs, usually under 50m. Perennial. Flowering period III-VI. Propagation method: seed germination in Petri dishes and transplanted in trays. Objective: Several true leaves, lignified and a plantable and healthy root mass. Material: - Sandpaper of very fine grit size (P150); - Distilled water; -	PROPAGATION PROTOCO Lotus azoricus FABACEA	E Secretaria Regional do Ambiente e Alterações Climáticas
YEAR 1 YEAR 2 YEAR 3 JAN FEB MAR APR MAY JUN JUL AUG SEP OCT NOV DEC JAN FEB MAR APR MAY JUN JUL AUG SEP OCT	Last Update: 24/05/2019 Ecology: Coastal cliffs, usually under 50m. Perennial. Flowering period III-VI. Propagation method: seed germination in Petri dishes and transplanted in trays. Objective: Several true leaves, lignified and a plantable and healthy root mass. Material: - Sandpaper of very fine grit size (P150); - Germination chamber; - Petri dishes; - Trays of germination 60cm3 (350x251x87 mm); - Distilled water; - Germination substrate. - Duration: 5 months.	Flowering Seed collection Seed conservation Establishment (Seeding, germ, emer, 1st leaves formation) Rapid growth (greenhouse) Rapid growth (outside the greenhouse) Hardening Transplantation to final location
 Seed collection: It should be done from May to July. Collect dark and dry pods before seed dispersion. If seeds are green, place them in a translucent container at direct sun light for 2 days. Seed conservation: The seeds are removed from the pods and dehydrated at 15°C and 15% humidity, for a minimum period of 1 month. Until propagation seeds are kept at 4°C. Germination pre-treatment: Prior to germination procedures, seeds are phisically scarified with sandpaper for seed coat sharpening. <u>Caution</u>: do not scarify the seeds for too long, it may detroy the seeds. Seeds are well scarified if swelled before 20 minutes immersed in water. Germination: 85%; Germination is started in Petri dishes with moisturized filter papper at 20°C/10°C and 16h of ligth per day (germination chamber). Seeding: Once the cotyledons are formed, transfer immediately the seedigns to germination substrate. Substrate: Germination substrate. Contrainer: Trays of germination 60cm³ (250x251x87 mm). 	YEAR 1 YEAR 2 YEAR 3 JAN FEB MAR APR MAY JUN JUL AUG SEP OCT NOV DEC JAN FEB MAR APR MAY JUN JUL AUG SEP OCT NOV DEC JAN FEB MAR APR MAY JUN JUL	
Substrate: Germination substrate.	 Seed collection: It should be done from May to July. Collect dark and dry pods before seed dispersion. If seeds are green, place them in a translucent container at direct sun ligth for 2 days. Seed conservation: The seeds are removed from the pods and dehydrated at 15°C and 15% humidity, for a minimum period of 1 month. Until propagation seeds are kept at 4°C. Germination pre-treatment: Prior to germination procedures, seeds are phisically scarified with sandpaper for seed coat sharpening. <u>Caution</u>: do not scarify the seeds for too long, it may detroy the seeds. Seeds are well scarified if swelled before 20 minutes immersed in water. Germination: 85%; Germination is started in Petri dishes with moisturized filter papper at 20°C/10°C and 16h of ligth per day (germination chamber). Seeding: Once the cotyledons are formed, transfer immediately the seedigns to germination substate. 	 Place: Trays inside the greenhouse. Establishment: Trays inside the greenhouse with frequent but not excessive irrigation. 2x10 minutes period/day. Cut irrigation for 2 days if brown algae start to apear. Not shaded place. Rapid growth: Inside the greenhouse: Plants shall be kept in the greenhouse for about one month, maintaining the sprinkler irrigation. Plants do not need to be always moist. Outside the greenhouse: plants with about 5-10 cm are taken to a shaded area outside; the irrigation frequency is reduced to once a day. Duration: 1/2 months. Hardening: Plants are taken to an area with less shade. Duration: 2/3 months. Fertilizing: During the hardening phase, apply K-enriched liquid fertilizer (5-8-10) every 15 days. Phytosanitary treatments: In May apply preventative treatment against the <i>Tipula</i> sp. larvae.
	Substrate: Germination substrate.	

References: Pereira *et al.* (2012). 'Rapid and effective germination methods to overcome primary seed dormancy in several Azorean endemic species'. *Acta Horticulturae*, 938: 77 - 84. Freitas, CF (2016). *A conservação* ex situ *das plantas autóctones dos Açores no Jardim Botânico do Faial*. Master Thesis.



LIFE VIDALIA has received funded from the LIFE Programme of the European Union